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Title: Cerebrospinal Fluid Tau and Phosphotau As Biomarkers in Alzheimer's Disease Diagnosis

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#### **ABSTRACT**

Dementia is a progressive disorder that leads to memory loss and cognition impairment and affects daily function. Alzheimer disease (AD) is the main cause of dementia that characterized by loss of memory and cognition. AD pathologically is demonstrated by neuronal atrophy, synapse loss and the unusual reposition of amyloid- $\beta$  protein (A $\beta$ ) as senile plaques and hyperphosphorylated tau protein as neurofibrillary tangles (NFT). Tau is a microtubule associated protein mostly expressed in neurons. Site-specific phosphorylation regulates Tau function. In AD, the six adult tau isoforms are unusually phosphorylated that cause to form the paired helical filament. The different conditions of tau phosphorylation eventuate from the function of specific kinases and phosphatases. In recent years some biomarkers such as phospho tau 181, 199 and 231 had been assessed in cerebrospinal fluid (CSF) and blood and had been showed their elevation in AD. This article provides an overview of tau structure, functions, and its involvement in AD and its role as a CSF biomarker.

Key words: Alzheimer, Tau, Phospho tau, Dementia, CSF biomarker, NFT

## **INTRODUCTION**

Dementia also known senility is a progressive disturbance which causes memory loss and cognition impairment and affects daily functioning of patients[1]. Observations show a significant increasing in the number of demented individuals from 25 million in the year 2000 to 63 million in 2030[2]. Alzheimer disease (AD) is the main cause of dementia and illustrates significant social problems that mostly is treated symptomatically [3]. Patients are demonstrated with memory loss then consequently present, confusion, agitation, and behavioral disorders [4]. The diagnosis of AD depends on clinical and neuropathological evaluation [5]. Rapidly progression of this disorder levitates from the molecular pathology in the limbic and related cortexes[6]. In the cognitively normal brain, there is an age-related reduction in brain volume and weight, enlargement of ventricles, and loss of synapses and dendrites in selected areas[7]. But AD pathologically is demonstrated by neuronal atrophy, synapse loss and the unusual reposition of amyloid-β protein (Aβ) as senile plaques and hyperphosphorylated tau protein as neurofibrillary tangles (NFT)[8]. Senile plaques are made of A $\beta$ -protein and the gene encoding A $\beta$  is related to AD [9] while, microtubule associated protein tau (MAPT) the gene encoding tau protein is not genetically associated to AD[10]. Illustrations show NFT implicate medial temporal lobe structures (hippocampus and entorhinal cortex) and then progress to temporal, parietal and frontal lobe[11].

#### Tau

Tau is a microtubule associated protein mostly expressed in neurons [12]. The human tau gene is located over 100 kb on the long arm of chromosome 17 at band position 17q21 and contains 16 exons [13]. Tau protein stabilizes microtubules and authorizes neurite' extension and stabilization [14].(FIG1) .The protein expresses in the axons of the central nervous system and was seen mainly in six isoforms with different functions which array from 352 to 441 amino acid and produced by alternative splicing. Abnormal deposition misprocessed Tau is an important pathway Alzheimer's disease. Tau has two ways of controlling microtubule stability: <u>isoforms</u> and <u>phosphorylation</u>. [15, 16]. These isoforms vary from each other by the attendance of either three- (3R) or four-repeat regions (4R) in the carboxy-terminal and the absence or presence of one or two inserts (29 or 58 amino acids) in the amino-terminal [17].

## Tau kinases and phosphatases

Protein phosphorylation is one of the major molecular mechanism which regulates protein function in reaction to extracellular stimulation of the nervous system[18]. Phosphorylation disport indispensable roles in signaling pathways and cellular processes like transcription, apoptosis, intercellular communication, and neuronal and immunological functions[19]. (FIG2). Observation

show site-specific phosphorylation of Tau regulates its function [20]. In accordance with these results phosphorylation of a few specific sites of tau is essential in tau-microtubule interactions regulations[21]. In the other hand hyperphosphorylation of tau reduces the biological activity of tau [22].Illustrations show tau is transiently phosphorylated during development and during anesthesia and hypothermia [23]. Normal adult human brain tau contains 2-3 moles phosphate/mole of tau protein[23]. While in AD, brain tau is three to four-fold more hyperphosphorylated than the normal adult brain tau and in this hyperphosphorylated state is polymerized into paired helical filaments(PHF) [24, 25]. In AD, the six adult tau isoforms are unusually phosphorylated that cause to form the PHF, the main fibrous component of the neurofibrillary lesions[26]. The longest type of adult tau in the human brain (441 amino-acids) has 80 Serine or Threonine residues and 5 Tyrosine residues; therefore, approximately 20% of the molecule has the potential of phosphorylation[27]. Tau is phosphorylated at almost 45 sites in AD brains. Tau aggregations differ in phosphorylation and content of tau isoforms, which enable a molecular classification of tauopathies[28]. Many of the phosphorylation sites are on Serine-Proline and Threonine-Proline sites. But few of them are on non-Serine / Threonine-Proline motives [13]. The different conditions of tau phosphorylation eventuate from the function of specific kinases and phosphatases[29].

The main tau kinases are glycogen-synthase kinase-3β (GSK-3β), cyclin-dependent protein kinase 5 (cdk5), cAMP-dependent protein kinase (PKA), and stress-activated protein kinases[30]. Glycogen synthase kinase 3 (GSK3) is a serine/threonine protein kinase that involves in phosphorylation of serine and threonine amino acids [31]. GSK-3 is encoded by two genes, GSK-3 alpha (GSK3A) and GSK-3 beta (33GSK3B)[32]. Phosphorylation of Thr231 have the main role in regulating tau-microtubule interactions and Thr231 is the brief site for GSK3β [33, 34]. Phosphorylation of Thr231 remarkedably reduces tau and microtubules binding [34]. Cdk5 is a member of the cyclin-dependent kinases (Cdks) that activated in post-mitotic neurons through the neuron-specific activator p35[35, 36]. Cdk5-p35 has the main role in brain development and physiological synaptic functions [37]. In pathologic states Cdk5 is hyperactivated by p25 [38], that is the N-terminal shortened form of p35[39]. There is a reposition of p25 in AD brains with high phosphorylation capability of Cdk5-p25 for tau 0in comparison with Cdk5-p35[40]. Between the 16 Ser/Thr-Pro sites in tau, Cdk5 phosphorylates 9–13 residue[41]. The in vitro investigations show Ser202, Thr205, Ser235, and Ser404 are the major residues and Thr153 and Thr212 are the

minor sites of phosphorylation[41]. phosphorylation at serine 422 in tau occurs early and related to cognitive decline in AD[42]. Elevated levels of t-tau and p-tau181 in CSF show degenerative processes in the cortical areas specially affected in Alzheimer disease. p-tau181 can be more significantly associated with neurodegenerative alternations in early AD[43] .The CSF/phosphotau199 levels in the AD are remarkedably high with 85.2 in sensitivity and specificity.So Csf phosphotau199 can be a new biomarker in diagnosis of AD [44].

Protein phosphatases( PP2A) is the major tau phosphatase in the brain[45]. All protein phosphatases, involving PP2A, have more broad substrate specificity than protein kinases[46]. PP2A activity is regulated by two endogenous protein inhibitors, I<sub>1</sub>PP2A and I<sub>2</sub>PP2A [47]. These two inhibitors may be disregulated in AD brain cause to PP2A inhibition [48]

## Tau as peripheral biomarkers

Tau first was described in cerebrospinal fluid (CSF) in 1993 [49]. Total Tau in CSF was first quantified by ELISA method that assesses all Tau isoforms [50]. The results illustrated that CSF total Tau(CSF-tTau) increases with an age-related pattern. Elevation in CSF-tau is observed in Alzheimer's .the same high CSF-tau concentrations may be seen before the onset of clinical dementia.. [51]. Illustrations demonstrated that elevations of CSF-tTau in AD are 2-3 times higher as compared to control group[50].CSF-tTau increase due to Tau exudation from neurons into CSF and is a sign of neuronal damage severity [52]. In fact, There is a growing body of evidence implicating that in acute states like stroke or Creutzfeld Jacob disease (CJD), a marked elevation in CSF-tTau was seen[53, 54]. Bearing this in mind phosphorylated tau (p-Tau) is one of the important biomarkers in early diagnosis of AD. [55]. In the other hand elevation of CSF-pTau as compared to controls has found in all ELISA assays [56]. Observations show pTau181, pTau199, and pTau231 assays differ in AD from normal aging[57]. Evidences implicate the specificity of pTau is more than total Tau in differentiating various conditions[58]. Diversity in phosphoepitopes is the main factor in their potential to forejudge AD from other neurodegenerative [59]. In agreement with this conclusion, The best difference between AD and Frontotemporal Dementia (FTD) is pTau231 and between AD and Dementia with Lewy Bodies (DLB) is pTau181[60, 61]. Several studies show that high CSF-tTau and CSF-pTau levels are seen in early AD and the performance of these biomarkers in AD with MMSE (Mini-Mental State Examination) scores above 23-25 is similar to advanced AD[62]. As well as, in MCI progressing to AD during clinical

follow-up, the biomarkers are changed with sensitivities equivalent to AD dementia [63]. PTau at various epitopes including Thr181, Thr231 and Ser 199 is an authentic predictor of the conversion from MCI to AD, with a sensitivity of 66- 100% and a specificity of 66-78%[52]. In addition elevation of tTau/pTau ratio is a valuable marker in differential early diagnosis of dementia[52]. P-tau231 and t-tau levels are higher in patients with MCI than in control. Observations show CSF p-tau231 levels increase in MCI [64]. In addition High p-tau231 levels are related to the point loss in MMSE scores MCI [65]. CSF p-tau231 elevation has a prediction role in cognitive decline in patients with MCI[66]{Randall, 2013 #3058}. Other studies point to p-tau 181 concentrations as a introducer of the clinical symptoms of AD [66]. Several studies have provided evidence that CSF t-tau and p-tau181p are related to neuronal and axonal damage[67]. In progressive MCI CSF p-tau181 and t-tau elevation are related to hippocampal atrophy[68]. P-tau181, and t-tau are related to specific model in brain structure changes in AD [69]. In table 1 there are some measures of various p-tau and t-tau in different studies in AD or MCI.

## **DISCUSION**

Alzheimer disease (AD) is the main cause of dementia and illustrates significant social problems that mostly is treated symptomatically [3]. In the cognitively normal brain, there is an age-related reduction in brain volume and weight, enlargement of ventricles, and loss of synapses and dendrites in selected areas[7]. AD pathologically is demonstrated by neuronal atrophy, synapse loss and the unusual reposition of amyloid- $\beta$  protein (A $\beta$ ) as senile plaques and hyperphosphorylated tau protein as neurofibrillary tangles (NFT). As a matter of fact Tau is a microtubule associated protein mostly expressed in neurons. These proteins are found mostly in neurons compared to non-neuronal cells. One of tau's main functions is to modulate the stability of axonal microtubules. [8]. CSF-tTau increases due to Tau exudation from neurons into CSF and is a sign of neuronal damage severity [52]. There is evidence implicating that the specificity of pTau is more than total Tau in differentialing various conditions [58]. In addition elevation of tTau/pTau ratio is a valuable marker in differential early diagnosis of dementia [52]. In recent years serum or CSF tau protein as a most important biomarker have been used in diagnosis of AD. CSF

total tau and phosphotau was the aim of some studies. But because of LP invasiveness researches is low number. However the results of these studies are different, that can be due to the selection of patients in different stages of the disease or using different method. So, more studies with large number of patients in different stages and accurate methods are necessary.

# **Abrevations**

AD: Alzheimer disease

Aβ: Amyloid-β protein

NFT: Neurofibrillary tangles

MPT: Microtubule associated protein tau

GSK3: Glycogen synthase kinase 3

CDKS: Cyclin-dependent kinases

PP2A: Protein phosphatases

P-TAU: Phospho TAU

MCI: Mild cognitive impairment

CSF: Cerebro spinal fluid

FTD: Frontotemporal Dementia

DLB: Dementia with Lewy Bodies

CJD: Creutzfeld Jacob disease

#### REFRENCES

- 1. Chiang H-Y. Dementia risk and medical cost assessment model for patients with stroke. 2016.
- 2. Wimo A, Winblad B, Aguero-Torres H, von Strauss E. The magnitude of dementia occurrence in the world. Alzheimer Disease & Associated Disorders. 2003;17(2):63-7.
- 3. Sadigh-Eteghad S, Majdi A, Farhoudi M, Talebi M, Mahmoudi J. Different patterns of brain activation in normal aging and Alzheimer's disease from cognitional sight: meta analysis using activation likelihood estimation. Journal of the neurological sciences. 2014;343(1):159-66.
- 4. Bekris LM, Yu C-E, Bird TD, Tsuang DW. Review article: genetics of Alzheimer disease. Journal of geriatric psychiatry and neurology. 2010;23(4):213-27.
- 5. Bird TD. Alzheimer disease overview. 2014.
- 6. Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. Physiological reviews. 2001;81(2):741-66.
- 7. Armstrong RA. What causes Alzheimer's disease. Folia Neuropathol. 2013;51(3):169-88.
- 8. Holtzman DM, Morris JC, Goate AM. Alzheimer's disease: the challenge of the second century. Science translational medicine. 2011;3(77):77sr1-sr1.
- 9. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science. 2002;297(5580):353-6.
- 10. Wolfe MS. The role of tau in neurodegenerative diseases and its potential as a therapeutic target. Scientifica. 2012;2012.
- 11. Karantzoulis S, Galvin JE. Distinguishing Alzheimer's disease from other major forms of dementia. Expert review of neurotherapeutics. 2014.
- 12. Ingram EM, Spillantini MG. Tau gene mutations: dissecting the pathogenesis of FTDP-17. Trends in molecular medicine. 2002;8(12):555-62.
- 13. Buée L, Bussiere T, Buée-Scherrer V, Delacourte A, Hof PR. Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. Brain Research Reviews. 2000;33(1):95-130.
- 14. Liu CwA, Lee G, Jay DG. Tau is required for neurite outgrowth and growth cone motility of chick sensory neurons. Cell motility and the cytoskeleton. 1999;43(3):232-42.
- 15. Mandelkow E-M, Mandelkow E. Biochemistry and cell biology of tau protein in neurofibrillary degeneration. Cold Spring Harbor perspectives in medicine. 2012;2(7):a006247.
- 16. Goedert M, Jakes R. Expression of separate isoforms of human tau protein: correlation with the tau pattern in brain and effects on tubulin polymerization. The EMBO journal. 1990;9(13):4225.
- 17. Sergeant N, Delacourte A, Buée L. Tau protein as a differential biomarker of tauopathies. Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease. 2005;1739(2):179-97.

- 18. Nestler EJ, Greengard P. Protein phosphorylation is of fundamental importance in biological Regulation. 1999.
- 19. Johnson LN. The regulation of protein phosphorylation. Biochemical Society Transactions. 2009;37(4):627-41.
- 20. Stoothoff WH, Johnson GV. Tau phosphorylation: physiological and pathological consequences. Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease. 2005;1739(2):280-97.
- 21. Billingsley ML, Kincaid RL. Regulated phosphorylation and dephosphorylation of tau protein: effects on microtubule interaction, intracellular trafficking and neurodegeneration. Biochemical Journal. 1997;323(3):577-91.
- 22. Wang J-Z, Xia Y-Y, Grundke-Iqbal I, Iqbal K. Abnormal hyperphosphorylation of tau: sites, regulation, and molecular mechanism of neurofibrillary degeneration. Journal of Alzheimer's Disease. 2013;33(s1):S123-S39.
- 23. Iqbal K, Liu F, Gong C-X, Grundke-Iqbal I. Tau in Alzheimer disease and related tauopathies. Current Alzheimer Research. 2010;7(8):656-64.
- 24. Alonso AdC, Zaidi T, Novak M, Grundke-Iqbal I, Iqbal K. Hyperphosphorylation induces self-assembly of  $\tau$  into tangles of paired helical filaments/straight filaments. Proceedings of the National Academy of Sciences. 2001;98(12):6923-8.
- 25. Rossi D. Analysis of Reelin function in the molecular mechanisms underlying Alzheimer's disease: Universitat de Barcelona; 2013.
- 26. Goedert M, Jakes R, Crowther R, Six J, Lübke U, Vandermeeren M, et al. The abnormal phosphorylation of tau protein at Ser-202 in Alzheimer disease recapitulates phosphorylation during development. Proceedings of the National Academy of Sciences. 1993;90(11):5066-70.
- 27. Johnson GV, Stoothoff WH. Tau phosphorylation in neuronal cell function and dysfunction. Journal of cell science. 2004;117(24):5721-9.
- 28. Williams D. Tauopathies: Classification and clinical update on neurodegenerative diseases associated with microtubule-associated protein tau. Internal medicine journal. 2006;36(10):652-60.
- 29. Gong C-X, Lidsky T, Wegiel J, Zuck L, Grundke-Iqbal I, Iqbal K. Phosphorylation of microtubule-associated protein tau is regulated by protein phosphatase 2A in mammalian brain implications for neurofibrillary degeneration in Alzheimer's disease. Journal of Biological Chemistry. 2000;275(8):5535-44.
- 30. Dolan PJ, Johnson GV. The role of tau kinases in Alzheimer's disease. Current opinion in drug discovery & development. 2010;13(5):595.
- 31. Embi N, RYLATT DB, COHEN P. Glycogen Synthase Kinase-3 from Rabbit Skeletal Muscle. European Journal of biochemistry. 1980;107(2):519-27.
- 32. Hoeflich KP, Luo J, Rubie EA, Tsao M-S, Jin O, Woodgett JR. Requirement for glycogen synthase kinase-3β in cell survival and NF-κB activation. Nature. 2000;406(6791):86-90.
- 33. Sottejeau Y, Bretteville A, Cantrelle F-X, Malmanche N, Demiaute F, Mendes T, et al. Tau phosphorylation regulates the interaction between BIN1's SH3 domain and Tau's proline-rich domain. Acta neuropathologica communications. 2015;3(1):1.
- 34. Cho JH, Johnson GV. Primed phosphorylation of tau at Thr231 by glycogen synthase kinase  $3\beta$  (GSK3 $\beta$ ) plays a critical role in regulating tau's ability to bind and stabilize microtubules. Journal of neurochemistry. 2004;88(2):349-58.
- 35. Malumbres M, Barbacid M. Mammalian cyclin-dependent kinases. Trends in biochemical sciences. 2005;30(11):630-41.

- 36. Patrick GN, Zhou P, Kwon YT, Howley PM, Tsai L-H. p35, the neuronal-specific activator of cyclin-dependent kinase 5 (Cdk5) is degraded by the ubiquitin-proteasome pathway. Journal of Biological Chemistry. 1998; 273(37):24057-64.
- 37. Nikolic M, Dudek H, Kwon YT, Ramos Y, Tsai L-H. The cdk5/p35 kinase is essential for neurite outgrowth during neuronal differentiation. Genes & development. 1996;10(7):816-25.
- 38. Su SC, Tsai L-H. Cyclin-dependent kinases in brain development and disease. Annual review of cell and developmental biology. 2011; 27:465-91.
- 39. Mishiba T, Tanaka M, Mita N, He X, Sasamoto K, Itohara S, et al. Cdk5/p35 functions as a crucial regulator of spatial learning and memory. Molecular brain. 2014;7(1):1.
- 40. Patrick GN, Zukerberg L, Nikolic M, de La Monte S, Dikkes P, Tsai L-H. Conversion of p35 to p25 deregulates Cdk5 activity and promotes neurodegeneration. Nature. 1999;402(6762):615-22.
- 41. Kimura T, Ishiguro K, Hisanaga S-i. Physiological and pathological phosphorylation of tau by Cdk5. Frontiers in molecular neuroscience. 2014;7:65.
- 42. Kanaan NM, Cox K, Alvarez VE, Stein TD, Poncil S, McKee AC. Characterization of Early Pathological Tau Conformations and Phosphorylation in Chronic Traumatic Encephalopathy. Journal of Neuropathology & Experimental Neurology. 2015:nlv001.
- 43. Thomann PA, Kaiser E, Schönknecht P. Association of total tau and phosphorylated tau 181 protein levels in cerebrospinal fluid with cerebral atrophy in mild cognitive impairment and Alzheimer disease. Journal of psychiatry & neuroscience: JPN. 2009;34(2):136.
- 44. Itoh N, Arai H, Urakami K, Ishiguro K, Ohno H, Hampel H, et al. Large-scale, multicenter study of cerebrospinal fluid tau protein phosphorylated at serine 199 for the antemortem diagnosis of Alzheimer's disease. Annals of neurology. 2001;50(2):150-6.
- 45. Sontag J-M, Nunbhakdi-Craig V, White CL, Halpain S, Sontag E. The protein phosphatase PP2A/Bα binds to the microtubule-associated proteins tau and MAP2 at a motif also recognized by the kinase Fyn implications for tauopathies. Journal of Biological Chemistry. 2012;287(18):14984-93.
- 46. Shi Y. Serine/threonine phosphatases: mechanism through structure. Cell. 2009;139(3):468-84.
- 47. Liu C, Götz J. How it all started: tau and protein phosphatase 2A. Journal of Alzheimer's Disease. 2013;37(3):483-94.
- 48. Gong C-X, Iqbal K. Hyperphosphorylation of microtubule-associated protein tau: a promising therapeutic target for Alzheimer disease. Current medicinal chemistry. 2008;15(23):2321-8.
- 49. Yamamori H, Khatoon S, Grundke-Iqbal I, Blennow K, Ewers M, Hampel H, et al. Tau in cerebrospinal fluid: a sensitive sandwich enzyme-linked immunosorbent assay using tyramide signal amplification. Neuroscience letters. 2007;418(2):186-9.
- 50. Blennow K. Cerebrospinal fluid protein biomarkers for Alzheimer's disease. NeuroRx. 2004;1(2):213-25.
- 51. Buerger K, Zinkowski R, Teipel SJ, Arai H, DeBernardis J, Kerkman D, et al. Differentiation of geriatric major depression from Alzheimer's disease with CSF tau protein phosphorylated at threonine 231. American Journal of Psychiatry. 2003.
- 52. Schraen-Maschke S, Sergeant N, Dhaenens C-M, Bombois S, Deramecourt V, Caillet-Boudin M-L, et al. Tau as a biomarker of neurodegenerative diseases. 2008.

- 53. Blennow K, Lind B, Andersson E, Andreasen N. [CSF-analyses in clinical diagnosis of Creutzfeldt-Jakob disease. A literature review and three cases from routine clinical practice]. Lakartidningen. 2001;98(20):2446-51.
- 54. Sjögren M, Vanderstichele H, Ågren H, Zachrisson O, Edsbagge M, Wikkelsø C, et al. Tau and Aβ42 in cerebrospinal fluid from healthy adults 21–93 years of age: establishment of reference values. Clinical chemistry. 2001;47(10):1776-81.
- 55. Kandimalla RJ, Prabhakar S, Wani WY, Kaushal A, Gupta N, Sharma DR, et al. CSF p-Tau levels in the prediction of Alzheimer's disease. Biology open. 2013:BIO20135447.
- 56. Hampel H, Blennow K. CSF tau and-amyloid as biomarkers for mild cognitive impairment. Dialogues Clin Neurosci. 2004;6:379-90.
- 57. Hampel H, Buerger K, Zinkowski R, Teipel SJ, Goernitz A, Andreasen N, et al. Measurement of Phosphorylated Tau Epitopes in the Differential Diagnosisof Alzheimer Disease: A Comparative Cerebrospinal Fluid Study. Archives of general psychiatry. 2004;61(1):95-102.
- 58. Galasko D. Cerebrospinal fluid biomarkers in Alzheimer disease: a fractional improvement? Archives of neurology. 2003;60(9):1195-6.
- 59. Bouwman FH. CSF biomarkers in dementia: Longitudinal aspects and combination with MRI: Amsterdam: Vrije Universiteit; 2008.
- 60. Wiltfang J, Lewczuk P, Riederer P, Grünblatt E, Hock C, Scheltens P, et al. Consensus Paper of the WFSBP Task Force on Biological Markers of Dementia: the role of CSF and blood analysis in the early and differential diagnosis of dementia (World J Biol Psychiatry. 2005; 6 (2): 69-84. Review). Archives of Clinical Psychiatry (São Paulo). 2009;36:1-16.
- 61. Mukaetova-Ladinska EB, Monteith R, Perry EK. Cerebrospinal fluid biomarkers for dementia with lewy bodies. International Journal of Alzheimer's Disease. 2010;2010.
- 62. Andreasen N, Blennow K. CSF biomarkers for mild cognitive impairment and early Alzheimer's disease. Clinical neurology and neurosurgery. 2005;107(3):165-73.
- 63. Andreasen N, Vanmechelen E, Vanderstichele H, Davidsson P, Blennow K. Cerebrospinal fluid levels of total-tau, phospho-tau and Aβ42 predicts development of Alzheimer's disease in patients with mild cognitive impairment. Acta Neurologica Scandinavica. 2003;107(s179):47-51.
- 64. Hampel H, Broich K. Enrichment of MCI and early Alzheimer's disease treatment trials using neurochemical & imaging candidate biomarkers. JNHA-The Journal of Nutrition, Health and Aging. 2009;13(4):373-5.
- 65. Kerkman D, DeBernardis J, Zinkowski R, Hampel H. Method for predicting whether subjects with mild cognitive impairment (MCI) will develop Alzheimer's Disease. Google Patents; 2003.
- 66. Fjell A, Walhovd K, Amlien I, Bjørnerud A, Reinvang I, Gjerstad L, et al. Morphometric changes in the episodic memory network and tau pathologic features correlate with memory performance in patients with mild cognitive impairment. American Journal of Neuroradiology. 2008;29(6):1183-9.
- 67. Clark CM, Xie S, Chittams J, Ewbank D, Peskind E, Galasko D, et al. Cerebrospinal fluid tau and  $\beta$ -amyloid: how well do these biomarkers reflect autopsy-confirmed dementia diagnoses? Archives of neurology. 2003;60(12):1696-702.
- 68. Henneman W, Vrenken H, Barnes J, Sluimer I, Verwey N, Blankenstein M, et al. Baseline CSF p-tau levels independently predict progression of hippocampal atrophy in Alzheimer disease. Neurology. 2009;73(12):935-40.

- 69. Tosun D, Schuff N, Truran-Sacrey D, Shaw LM, Trojanowski JQ, Aisen P, et al. Relations between brain tissue loss, CSF biomarkers, and the ApoE genetic profile: a longitudinal MRI study. Neurobiology of aging. 2010;31(8):1340-54.
- 70. De Jong D, Jansen RW, Kremer BP, Verbeek MM. Cerebrospinal fluid amyloid  $\beta$ 42/phosphorylated tau ratio discriminates between Alzheimer's disease and vascular dementia. The Journals of Gerontology Series A: Biological Sciences and Medical Sciences. 2006;61(7):755-8.
- 71. Monge-Argilés JA, Muñoz-Ruiz C, Sánchez-Payá J, Gasparini Berenguer R, Blanco Cantó ME, Leiva-Santana C. Comparison of two analytical platforms for CSF biomarkers of Alzheimer's disease. BioMed research international. 2014;2014.
- 72. Vandermeeren M, Mercken M, Vanmechelen E, Six J, Voorde A, Martin JJ, et al. Detection of Proteins in Normal and Alzheimer's Disease Cerebrospinal Fluid with a Sensitive Sandwich Enzyme-Linked Immunosorbent Assay. Journal of neurochemistry. 1993;61(5):1828-34.
- 73. Kaerst L, Kuhlmann A, Wedekind D, Stoeck K, Lange P, Zerr I. Cerebrospinal fluid biomarkers in Alzheimer's disease, vascular dementia and ischemic stroke patients: a critical analysis. Journal of neurology. 2013;260(11):2722-7.
- 74. Boban M, Grbić K, Mladinov M, R Hof P, Süßmair C, Ackl N, et al. Cerebrospinal fluid markers in differential diagnosis of Alzheimer's disease and vascular dementia. Collegium antropologicum. 2008;32(1):31-6.
- 75. Spiegel J, Pirraglia E, Osorio RS, Glodzik L, Li Y, Tsui W, et al. Greater Specificity for Cerebrospinal Fluid P-tau231 over P-tau181 in the Differentiation of Healthy Controls from Alzheimer's Disease. Journal of Alzheimer's Disease. 2015;49(1):93-100.

Table1: csf tau and phosphotau levels in MCI and AD

Reference	P-tau 181	p-tau231	p-tau199	t-tau
[57, 70]	103 pg/ml CSF	667.5 pg/ml csf	1.7 fmol/ml csf	631 pg/ml CSF
[71, 72]	86.71 pg/ml		20 pg/ml CSF-	572.73 pg/ml
	CSF-AD		AD	CSF
[73, 74]	339 pg/ml		58.5 pg/ml CSF-	70pg/ml (CSF-
	(CSF-AD)		AD	AD)
	200.5 pg/ml			57.5pg/ml(CSF-
	(CSF-MCI)			MCI)
[75]	94.8 pg/ml CSF-	64.4 pg/ml CSF-		
	AD	AD		

Abbreviations: AD=Alzheimer disease; MCI= mild cognitive impairment; CSF = cerebrospinal fluid

Fig 1: tau and microtubule interaction



Fig 2: protein phosphorylation with kinases and phosphatases



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